**ISAC Council Statement – Mariela Bollati-Fogolín**

I am honored to stand for election to ISAC Council as Councilor for Basic Research, bringing over **20 years of experience** bridging fundamental biological discovery and Shared Resource Laboratory (SRL) management, I bring a unique perspective on how technology empowers science. I currently serve as Head of the Cell Biology Unit at Institut Pasteur de Montevideo in Uruguay.

My experience as Chair of CYTO Women and as a member of the Live Education Delivery Committee has reinforced my belief that inclusive leadership can transform our Society. If elected, I will champion **global equity**, **expand multilingual education**, and **create structured pathways for Emerging Leaders** to actively shape ISAC's future.

I was privileged to receive the **2025 Howard Shapiro Award** for contributions to education in resource- limited settings. A central focus of my work has been to help establish Latin America as a hub for excellence in cytometry by organizing workshops and training programs across the region. In one recent initiative, we secured full funding to bring **25 students from 12 Latin American** countries to our Institute in Uruguay, all on full scholarships. Seeing them learn, interact, build lasting collaborations and lay the foundation for a new generation of cytometrists across Latin America.

I will collaborate with the other Council members and the Executive Committee on the following priorities in alignment with ISAC's goals:

**Advance equitable global access and expand ISAC's global reach (Goal 1)**

- Cytometry is increasingly global, with innovation extending beyond traditional hubs. ISAC should strengthen connections among emerging communities, building on models like CYTO Connect. I will strengthen regional partnerships with ISAC Associated Societies to deliver sustainable, high-quality training worldwide.
- I will work to bring expertise directly to geographically or scientifically isolated communities through hybrid educational models.

**Democratization and Diversity of Leadership (Goal 2)**

- I will advocate for structured pathways that enable early-career investigators, SRL professionals, innovators and scientists from emerging regions to engage in ISAC volunteer service and governance.
- I strongly support the continued growth of CYTO Women initiatives, including mentoring and sponsorship programs to advance leadership pathways.

**Enhance ISAC's global scientific presence and identity by empowering emerging leaders (Goals 3&4)**

- I will work to position ISAC Emerging Leaders and Scholars as ambassadors of the Society at interdisciplinary conferences, regional meetings, and partner scientific events.
- Creating Emerging Leader exchange programs, increasing visibility, and fostering cross-disciplinary dialogue will strengthen ISAC's global scientific presence while cultivating the next generation of cytometry leaders.
- I am committed to reinforcing member trust by clearly communicating strategic priorities and the measurable impact of our work.

**Educational Excellence and Multilingual Access (Goal 5)**

- I will continue to support multilingual initiatives, encouraging the delivery of courses and materials in diverse linguistic contexts to lower barriers to participation. Most of our Latin American workshops are already bilingual, presented in English and Spanish. We have translated ISAC's recent "Basics of Flow Cytometry" E-course by Zosia Maciorowski into Spanish, soon to be available on the ISAC YouTube channel, while the Mandarin version is already online.
- I support the integration of emerging technologies, reproducibility standards, and data science into CYTO U and other ISAC programming.

**ISAC Council Statement – Mariela Bollati-Fogolín**

**Closing Statement**

My deep-rooted involvement across multiple ISAC committees has given me the insight to navigate the Society's inner workings and turn strategic goals into reality. I believe sustainable growth requires inclusive leadership that enriches decision-making through diverse representation. Having led multidisciplinary teams and built international collaborations, I am prepared to contribute responsibly to ISAC's strategic vision and ensure we remain a diverse, innovative, and truly global cytometry community.

Qualifications

**ISAC Engagement:**

- Howard Shapiro Award 2025, for contributions to the field of cytometry and global health.
- 8 years as an active ISAC member
- Chair of ISAC CYTO Women Committee (4 years)
- Member of the CYTO organizing committee (2022 to 2025)
- Member of the CYTO program committee (2021, 2026)
- Member of ISAC Live Education Delivery Sub/committee of the Education Committee (4 years)
- Member of ISAC Instrument for Science (I4S) Task Force (4 years)

**Regional and International Leadership:**

- 4 years (two terms) on the GRCF (Uruguayan Argentine Flow Cytometry Society) Council
- GRCF representative on the ISAC Associated Societies Committee (2 years)

**Education and Training**

- 20 years enabling use of flow cytometry by scientists in Uruguay and Latin America
- 20 years developing and delivering flow cytometry training courses across Latin America, including Argentina, Brazil, Chile, Mexico and Uruguay.
- Extensive experience in flow cytometry education and training, including organizing workshops, symposia, preparing budgets and securing local, Latin American and international funding.
- Teaching faculty and organizer for ISAC Live Education workshops in Latin America

Biographical Sketch:

Prior to leading the Cell Biology Unit (CBU) at the Institut Pasteur de Montevideo, I obtained my degree in Biochemistry and my PhD in Biological Sciences from the Universidad Nacional del Litoral (Argentina). I subsequently completed postdoctoral training at the Helmholtz Centre for Infection Research in Germany, further specializing in infection biology and advanced cellular and molecular approaches. This foundation of international experience strengthened my scientific independence, expanded my collaborative network, and provided the basis for the leadership and technological development I later established within the CBU.

I have served as Head of the Cell Biology Unit (CBU) since 2006, integrating biochemistry and molecular biology with advanced experimental models to advance our understanding of complex cellular processes. For two decades, I have led the development of a state-of-the-art cytometry and advanced cell culture platform that has become a national and regional reference in Latin America. Managing the CBU Shared Resource Laboratory and its staff, I oversee scientific and strategic operations while providing researchers, students, and industry partners with access to advanced cytometry technologies and specialized expertise to advance their scientific and translational goals.

My research focuses on the development and application of 2D and 3D cell reporter models to address key pathophysiological processes, including inflammation and infectious diseases. I have authored more than **80 peer-reviewed publications** and trained and mentored 10 PhD and Master's students, hosting more than 20 international fellows primarily from Latin America and Europe, contributing to sustained regional capacity building.

I have led collaborative academia/industry projects and served as scientific consultant. Since 2021 I have served as the Uruguayan Director of the Centro Latinoamericano de Biotecnología (CABBIO), strengthening regional biotechnology integration. In recognition of my contributions to international scientific cooperation and cytometry, I have been honored with the 2025 RAICES Award and the 2025 ISAC Howard Shapiro Award.

MARIELA BOLLATI FOGOLÍN, Ph.D.

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## PROFESSIONAL PROFILE

Biochemist and PhD in Biological Sciences with extensive expertise in Life Sciences, leading the Cell Biology Unit at the Institut Pasteur de Montevideo since 2006. Her scientific background integrates biochemistry, molecular and cellular biology, and advanced experimental models. Under her direction, the unit has become a national and regional reference in flow cytometry and advanced cell culture technologies, providing scientific and technological support to researchers, students, and companies across Latin America.

## EDUCATION & TRAINING

**2002-2006:** **Postdoctoral researcher,** Helmholtz Centre for Infection Research, Germany.

**2002:** **PhD. in Biological Sciences**, Universidad Nacional del Litoral, Santa Fe, Argentina.

**1995:** Diploma in **Biochemistry**, School of Biochemistry and Biological Sciences, Universidad Nacional del Litoral, Santa Fe, Argentina.

## CURRENT & PREVIOUS POSITIONS

2006 – present: **Head of Cell Biology Unit**, Institut Pasteur de Montevideo. Montevideo, Uruguay.
2021 – present: **Uruguayan Director** of the Centro Latinoamericano de Biotecnología (CABBIO).
2024 – present: Lecturer in Diplomatura en Política y Gestión de la CyT en Iberoamérica - CLACSO-CIICTI.
2021 – 2024: Chair of the ISAC CYTO Women Taskforce.
2019: Visiting Professor at the Pontificia Universidad Católica de Valparaíso (Chile).
2011 – 2021: Professor of Cell Biology, School of Biotechnology, Ort University, Montevideo, Uruguay.
2002 – 2006: Post-Doctoral fellow, Experimental Immunology Group, Helmholtz Centre for Infection Research (HZI). Braunschweig. Germany.
2000 – 2002: Fellow Scientist, Cell Culture Technology Department (ZKT), German Research Centre for Biotechnology (GBF). Germany.
1998 – 2002: Doctoral fellow, from the Universidad Nacional del Litoral, Santa Fe, Argentina.
1995 – 2000: Research Assistant, Cell Culture Laboratory. School of Biochemistry and Biological Sciences, Universidad Nacional del Litoral, Santa Fe, Argentina.
1993 – 1995: Assistant of Experimental activities (General Chemistry, Physical Chemistry and Immunology), Universidad Nacional del Litoral, Santa Fe, Argentina.
1992-1993: Scholarship for undergraduate students, Universidad Nacional de Litoral, Santa Fe, Argentina.

## CURRENT & PREVIOUS GRANTS AWARDED (as Principal Investigator or co-Principal Investigator)

**2023-2025:** ACIP 532-22. Intestinal organoid models for studying host-parasite interaction in Chagas disease.

**2020-2023**: ANII (FMV_1_2019_1_156213). Mini-intestinos: una potente herramienta *in vitro* para el reemplazo de animales de experimentación.

**2020-2021**: ANII (PEC_3 158811). Detección de 16 parámetros celulares mediante la incorporación de dos láseres al citómetro Attune NxT.

**2016-2019**: ANII (ALI_1_2015_1_5084). Diseño y producción de nuevas variantes de la hormona folículo estimulante (FSH) para su empleo en especies de interés productivo.

**2016-2018**: ANII-CONICET (*MOV*_CO_2015_1_110054). Generación y caracterización de modelos in vitro para el estudio de perturbadores endócrinos.

**2014-2015**: ANII (EQC_2013_X_1_2). Separador celular de alta velocidad, multiparamétrico y bioseguro para su utilización en biomedicina y biotecnología.

**2013-2015**: ANII (FMV_2_2011_1_6046). Evaluación del riesgo por exposición a estrógenos ambientales antropogénicos en un modelo de ratones transgénicos Oct-4-GFP.

**2012-2013**: Cooperación internacional bilateral Uruguay-Argentina (DICyT y MEC). Monitoreo de compuestos naturales y sintéticos que modulen la actividad biológica de los interferones de tipo-I utilizando un nuevo ensayo gen reportero.

**2010-2012**: ANII (PR_FMV_2009_1_2617). Generación y caracterización de líneas celulares de importancia biotecnológica y biomédica para el monitoreo de cambios redox intracelulares en tiempo real e in situ.

**2010-2011**: ANII (PE_ALI_2009_1_1525). Desarrollo de un nuevo proceso de producción de vacunas virales para uso veterinario mediante el empleo de cultivos de alta densidad.

**GRANTS AWARDED FOR DELIVERING COURSES & WORKSHOPS**

**2024:** UNU-BIOLAC Organoids model: Basics to applications
**2024:** Welcome Connecting Science. Organoids model: Basics to applications
**2023:** UNU-BIOLAC Flow cytometry: from conventional to full spectrum training course
**2023:** ANII (VCT_X_2022_1_174766) Visiting Scientists for Flow Cytometry Training course.
**2021:** UNU-BIOLAC Webinar Series High content and high predictive cellular models for host-pathogen interactions studies, disease modeling and drug discovery
**2017:** ICGEB Course Grant. Cell and Animal Models for Drug Discovery
**2017:** Institut Pasteur International Network. Cell and Animal Models for Drug Discovery
**2017:** UNU-BIOLAC Course Grant. Cell and Animal Models for Drug Discovery
**2014:** ICGEB Grant. Flow cytometry and cell sorting in biotechnology and biomedicine research
**2014:** Institut Pasteur International Network. Flow cytometry and cell sorting in biotechnology and biomedicine research
**2014:** UNU-BIOLAC. Flow cytometry and cell sorting in biotechnology and biomedicine research
**2010:** ICGEB. Animal Cell Biotechnology: Product from cells, cells as product
**2010:** UNU-BIOLAC. Animal Cell Biotechnology: Product from cells, cells as product

**PUBLICATIONS**

**Peer-reviewed Manuscript** (H-Index: **27**, I-Index:**58** according to Google Scholar).

87. Sardi F, Elso O, Fiestas L, Perelmuter K, Benítez D, Corlatti A, Da Costa F, Lenz M, Schmidt T, **Bollati-Fogolín M**, Medeiros A, Sülsen V, Comini M. Insights into the Chemico-Biological Interactions of Natural Sesquiterpene Lactones with the Thiol-Redox Metabolism of Pathogenic Trypanosomatids. Accepted in ChemMedChem.
86. Bassetti L, Crispo M, **Bollati-Fogolin M**, Pantano S, Comini MA, Abreu C. Exploring Elastin-Like Polypeptide Tags and Mini-intein for Recombinant Protein Purification in Leishmania tarentolae. Protein Expr Purif. 2026 Feb 19:106902. doi: 10.1016/j.pep.2026.106902.
85. Cancela S, Sena F, Pagotto R, Francia ME, **Bollati-Fogolín M**. Protocol to enhance pre-sexual & sexual differentiation of Toxoplasma gondii using a retinal cells and intestinal organoid-derived monolayers. STAR Protoc. 2026 Feb 11;7(1):104367. doi: 10.1016/j.xpro.2026.104367.
84. Centomo AM, Díaz Vergara LI, Levit R, Perelmuter K, Bodoira RM, Moreno de LeBlanc A, LeBlanc JG, **Bollati-Fogolín M**, Maestri DM, Montenegro MA, Rossi Y. Functional evaluation of peanut skin phenolic extract and its microencapsulated form in oxidative stress and inflammation models. Discover Food (2026), Volume 6, article number 123.
83. Rodríguez-Basso AG, Prado HJ, Matulewicz MC, Perelmuter K, Pagotto R, Bach H, Gorzalczany SB, **Bollati-Fogolín M**. Chemical Profile and In Vitro Protective Effects of Minthostachys verticillata

(Griseb.) Epling Aqueous Extract in Intestinal Inflammatory Environments. Plants (Basel). 2025 Dec 25;15(1):69. doi: 10.3390/plants15010069.

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81. Cancela S, Sena F, Pagotto R, Crispo M, Francia ME, **Bollati-Fogolín M**. Enhancing pre-sexual and sexual differentiation of Toxoplasma gondii using retinal epithelial cells and intestinal organoids. Cell Rep. 2025 Oct 28;44(10):116451. doi: 10.1016/j.celrep.2025.116451
80. Piattoni CV, Abreu C, Machado MR, Pantano S, Comini MA, **Bollati-Fogolín M**. A red-shifted biosensor for intracellular detection of cAMP: the CUTieR the better. Biochem Biophys Res Commun. 2025 Apr 21;766:151831. doi: 10.1016/j.bbrc.2025.151831
79. Aversa-Marnai M, Perretta A, Céspedes P, Quartiani I, Conijeski D, **Bollati-Fogolin M**, Villarino A, Silva-Álvarez V, Ferreira AM. Chronic heat stress reshapes Russian sturgeon innate immune response to Aeromonas hydrophila challenge. Fish Shellfish Immunol. 2025 May;160:110219. doi: 10.1016/j.fsi.2025.110219.
78. Abud JE, Pagotto R, Galliani V , Teglia C , Culzoni J , **Bollati-Fogolín M** , Zenclussen ML, Rodríguez HA. In vitro blastocyst implantation and trophoblast migration are disrupted by the UV filter benzophenone-3 (BP3). Environmental Pollution, https://doi.org/10.1016/j.envpol.2024.123840.
77. Abreu C, Grunberg K, Bonilla M, Crispo M, Pantano S, Jaeschke J, Comini MA, **Bollati-Fogolín M.** Expression and Functional Characterization of Chimeric Recombinant Bovine Follicle-Stimulating Hormone Produced in Leishmania tarentolae. Microbial Biotechnology. doi: 10.1111/1751-7915.14444
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73. Fariña E, Daghero H, **Bollati-Fogolín M**, Boido M, Cantero J, Moncada-Basualto M, Olea-Azar C, Polticelli F, Paulino M (2023). Antioxidant capacity and NF-kB-mediated antiinflammatory 2 activity

of six Red Uruguayan grape pomaces. Molecules, 28(9):3909. doi: https://doi.org/10.3390/molecules28093909

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64. Arbildi P, Rodríguez-Camejo C, Perelmuter K, **Bollati-Fogolín M**, Sóñora C, Hernández A. Hypoxia and inflammation conditions differentially affect the expression of tissue transglutaminase spliced variants and functional properties of extravillous trophoblast cells. Am J Reprod Immunol. 2022. doi: 10.1111/aji.13534
63. Perelmuter K, Tiscornia I, Comini MA, **Bollati-Fogolín M**. Generation and characterization of stable redox-reporter mammalian cell lines of biotechnological relevance. Sensors (Basel). 2022. doi: https://doi.org/10.3390/s22041324.
62. Ruiz MC, Perelmulter K, Levín P, Romo AIB, Lemus L, **Fogolín MB**, León IE, Di Virgilio AL. Antiproliferative activity of two copper (II) complexes on colorectal cancer cell models: Impact on ROS production, apoptosis induction and NF-κB inhibition. Eur J Pharm Sci. 2022. doi: https://doi.org/10.1016/j.ejps.2021.106092
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5- **Bollati-Fogolín M**., Müller W. (2005) Virus free, cell-based assay for the quantification of murine type l Interferons. J. Immunol. Methods 306:169-175. doi: 10.1016/j.jim.2005.08.005 .

4- **Bollati-Fogolín M**., Forno G., Nimtz M., Conradt H., Etcheverrigaray M., Kratje R. (2005) Temperature Reduction in Cultures of hGM-CSF-expressing CHO Cells: Effect on Productivity and Product Quality. Biotechnol. Prog. 21(1): 17-21. doi: 10.1021/bp049825t .

3- **Fogolín M.B.,** Wagner R., Etcheverrigaray M., Kratje R. (2004) Impact of Temperature Reduction and Expression of Yeast Pyruvate Carboxylase on hGM-CSF-producing CHO cells. J. Biotechnol. 109 (1-2): 179-91. doi: 10.1016/j.jbiotec.2003.10.035

2- Forno G., **Bollati-Fogolín M**., Oggero Eberhardt M., Kratje R., Etcheverrigaray M., Conradt H., Nimtz M. (2004) N- and O-Glycosylation and Site Occupancy in Human Granulocyte-Macrophage Colony-Stimulating Factor Secreted by a Chinese Hamster Ovary Cell Line. Eur. J. Biochem. 271(5): 907-19. doi: 10.1111/j.1432-1033.2004.03993.x .

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**Book Chapters**

1- **Bollati-Fogolín M** and Comini M. (2008) Clonagem e expressão de proteinas heterologas em células animais. in: Tecnologia do Cultivo de Céluas Animais: de Biofármacos a Terapia Genica. Castilho, Moraes, Augusto (Eds), Published by Editora Roca Ltda, São Paulo, Brasil, 3: 40-76.

2- **Bollati-Fogolín M** and Comini M (2007), Cloning and expression of heterologous proteins in animal cells. in: Animal Cell Technology: From Biopharmaceuticals to Gene Therapy, Castilho, Moraes, Augusto, Butler (Eds), Published by Taylor & Francis Group. 3:39-73.

3- Ceaglio N, **Bollati-Fogolín M**, Oggero M, Etcheverrigaray M, Kratje R (2014).Ch 13: High cell density cultivation processes. In Animal Cell Biotechnology in Biologics Production, Eds Wagner R and Hauser H. De Gruyter. Berlín, Alemania (ISBN 3110278863, 9783110278866), 2014, pp. 427 – 454.

4- Perelmuter K, Arévalo AP, Crispo M, **Bollati-Fogolín M** (2020). Cap. 12: Determinación de la actividad biológica de glicoproteínas: modelos celulares y animales. En: Glicoproteínas Terapéuticas: diseño, expresión en células de mamíferos y análisis de sus glicanos. Editores: Forno G & Oggero M. Ediciones UNL. https://bibliotecavirtual.unl.edu.ar:8443/handle/11185/6277

**ORGANIZATION OF TRAINING COURSES**

- Organoids model: Basics to applications. Institut Pasteur de Montevideo, November 17-22, 2024. Supported by UNU-Biolac, IPMon, Wellcome Connecting Science. Organizers: **Bollati-Fogolín M**, Pagotto R.
- Pre-ALACI congress - Introductory flow cytometry course ISAC workshop. Universidad de Buenos Aires, Argentina. October 30 to November 2, 2024. Supported by ISAC, UNU-Biolac, UBA. Organizers: Quiroga F, González Polo V, Langer T**,** Billordo A, Collado S, Baz P, **Bollati-Fogolín M.**
- Flow Cytometry: From Conventional to Full Spectrum International Training Course (40 hours), Institut Pasteur de Montevideo, November 6-10, 2023. Supported by UNU-Biolac, ANII, ISAC, IPMon. Organizers: **Bollati-Fogolín M**, Telford W, Maciorowski Z, Céspedes MP y Perelmuter K.
- Fundamentos y aplicaciones de la citometría de flujo. Facultad de Bioquímica, Universidad Nacional del Litoral, Santa Fe, Argentina, September 18-22, 2023
- Flow Cytometry Workshop. Mode: virtual. March 15-17, 2022. Supported by ISAC. Organisers: **Mariela Bollati-Fogolín**, William Telford, Zosia M, Ma Paula Céspedes and Karen Perelmuter.
- Webinar series “High content and high predictive cellular models for host-pathogen interactions studies, disease modeling and drug discovery”, Mode: virtual. November, 2021. Supported by UNU Biolac. Organisers: **Mariela Bollati-Fogolín** and Romina Pagotto.
- Latin American School of Flow Cytometry. October 1-5, 2018. Institut Pasteur de Montevideo and Faculty of Medicine, Uruguay. Funded by ANII, IPMontevideo, GRCF, ISAC, Udelar, PEDECIBA, ProInBio. Course coordinators: **Bollati M**, Lens, Giordano,Blanco, Malusardi.
- Course “Alternative Methods to Lab Animals use”. May 21-25, 2018. Institut Pasteur de Montevideo, Uruguay. Funded by IPMontevideo, Premasur, LÓreal. Course coordinators: **Bollati M**, Crispo M, De Vecchi R.
- International Course “Cell and Animal Models for Drug Discovery”. October 16 to 27, 2017. Institut Pasteur de Montevideo. Funded by ICGEB, RIIP, UNU BIOLAC, ESACT and FOCEM. Course coordinators: **Bollati M**, Crispo M, Comini M, Alves P.
- Basic course in flow cytometry and its applications in research. February 22-26, 2016. Institut Pasteur de Montevideo, IIBCE, Uruguay. Funded by PEDECIBA. Coordinators of the course: Victoria S, Perelmuter K, **Bollati M**.
- International course “Flow cytometry and cell sorting in biotechnology and biomedicine research”. April 17-28, 2014. Institut Pasteur de Montevideo. Funded by RIIP, ICGEB, UNU-Biolac and TWAS. Course coordinators: **Bollati M**, Gröbe L, García JM.
- Course “Fundamentals and applications of flow cytometry”. October 7-18, 2013, Institut Pasteur de Montevideo, IIBCE, Uruguay. Sponsored by PEDECIBA and ProInBio. Course coordinators: Folle G, Porro V and **Bollati M**.
- IV Latin American Seminar on Cellular Technology. November 7-9, 2010 – Hotel Ermitage, Montevideo. Funded by ICGEB, UNU-Biolac. Organizing Committee: **Bollati M**, Kratje R.
- International course “Animal Cell Biotechnology: Product from cells, cells as product”, November 10-19, 2010, Institut Pasteur de Montevideo, Uruguay. Funded by ICGEB, UNU-Biolac. Organizing Committee: **Bollati M**, Kratje R.

**AWARDS**

- Recipient of the 2025 RAICES Award by the Secretaría de Innovación, Ciencia y Tecnología (Argentina). This award is intended for Argentine researchers based abroad who strengthen scientific cooperation with the country.
- Recipient of the 2025 **Howard Shapiro Award** by the International Society for Advancement of Cytometry (ISAC). This recognition highlights the outstanding contributions to the field of cytometry and global health.
- **SantanderW50 Uruguay**, selected in the 2025 edition of the women's leadership program aimed at strengthening the international impact of women with outstanding professional trajectories in Uruguay.
- Recipient of the 2023 **Premio de la Academia de Ciencias de Cuba** for the work *Nuevos aportes al conocimiento de la relación estructura-función en la actividad antitumoral del péptido CIGB-552*. Main authors: Julio R. Fernández Massó, Maribel Guerra Vallespi, Mariela Bollati Fogolín, Soledad Astrada Feijer, Hellen Daghero.

**CONTRIBUTION AS REVIEWER or ASSESOR**

Journals: PLoS ONE, J Biotechnol, Current Microbiology, Beneficial Microbes, Biochemical Engineering Journal, Molecules, Process Biochemistry, BMC Biotechnology, BioMed Research International, Journal of Immunological Sciences, Pharmaceutics.

Projects and fellowships: CONICYT (Chile), PICT (Argentina), ANII (Uruguay), National Science Centre (Poland), CABBIO (Brazil-Argentina-Uruguay), Austrian Academy of Sciences – L'Oréal Fellowship (Austria).

External Assessor: for a Professor of Practice/Director of Flow Cytometry Facility Position in the Biosciences Institute, Newcastle University. UK.

**STUDENTS SUPERVISION**

PhD students (4 finished, 1 present)

Master`s students (4 finished)

Undergraduate students (6 finished)

**OTHER RECOGNITIONS**

- Investigadora Nivel III del Sistema Nacional de Investigadores (SNI), Uruguay.
- Investigadora Grado 5 del Programa de Desarrollo de las Ciencias Básicas (PEDECIBA), Uruguay.
- Member of the board of directors del Grupo Rioplatense de Citometría de Flujo (GRCF), 2026-2027
- Since 2020 member of ISAC CYTO Women Taskforce and Chair from June 2021 to June 2024
- Founding member of Programa de Tecnología Molecular, Celular y Animal del IP-Montevideo
- Since 2022 member of ISAC Live Education Delivery and Instrument for Science (I4S) task forces.
- Founding member of Red Iberoamericana de Citometría de Flujo.
- Member of International Society for Advancement of Cytometry (ISAC), since 2018.
- Member of the European Society for Animal Cell Technology (ESACT), since 2003.
- Member of Grupo Rioplatense de Citometría de Flujo (GRCF)

**LANGUAGES**

- Spanish (native)
- English (advanced level)
- German (intermediate level)